

Prestige COVID-19 Lyophilised Nucleic Acid Kit (2-8°C)

CATALOGUE NUMBER	KIT SIZE (TESTS)	
PCRCOV1	50 Tests	

The COVID-19 Lyophilised Nucleic Acid Kit is an in vitro assay for the qualitative detection of nucleic acid of SARS-CoV-2 ORF1ab (open reading frame, ORF 1ab) and N (Nucleoprotein, N) gene in human respiratory tract secretion samples.

Coronaviruses are a large family of viruses that cause disease ranging from common cold symptoms to more severe pneumonia. They are enveloped, single strand RNA viruses. Coronaviruses are zoonotic, they can be transmitted from animals to humans. Existing examples include the Middle East Respiratory Virus (MER-CoV) and Severe Acute Respiratory Syndrome (SARS-CoV).

Reports of a novel coronavirus began in the Wuhan district of China in December 2019 and in January 2020 the World Health Organisation designated the new strain 2019-nCoV. Symptoms include high temperature, cough and breathing difficulties. In immunocompromised individuals symptoms can be more severe leading to pneumonia, severe acute respiratory syndrome or death.

Test Principle:

The kit combines the principles of real-time PCR and nucleic acid detection by use of fluorescent probes to quantify the SARS-CoV-2 ORF1ab and Nucleoprotein gene. The reagent includes a uracil-DNA glycosylase (UDG) enzyme to cleave the uracil glycosidic bond of uracil-containing PCR fragments as an efficient mechanism to reduce false positive results caused by PCR product contamination.

Reagents:

The kit comprises three tubes containing reagents of the following composition:

Tube	Contents	Volume	Quantity
1	1 Lyophilised Buffer Mix		3 vials
2	2 Reagent Solution		3 vials
3	3 Positive Control (Buffer PC)		1 vial
4 Negative Control (Buffer NC)		100 μΙ	1 vial

Instructions For Use sheet

Materials not provided:, PCR analyser, viral RNA extraction kit, Class II biological safety cabinet, vortex, microfuge nuclease free -centrifuge tubes and -PCR tubes/caps, micropipettes, sterile nuclease free pipette tips and sterile RNase- and nuclease-free distilled water.

Precautions:

For professional in vitro diagnostic use only.

Follow Good Laboratory Practice procedures where samples and kits are handled and treat the device and all samples as if potentially infectious. Follow local regulations for correct disposal of samples.

Wear protective clothing including laboratory coat, disposable gloves and safety glasses

when conducting the test.

Check the compatibility of the test reagents with the mode of action of the real-time PCR analyser. Compatible PCR analysers include Qiagen Rotor-Gene Q, Agilent Mx-3000p, Agilent Mx-3005p, ABI QuantStudio 3, Molarray MA-6000, Hongshi SLAN-96, Tianlong Gentier 96E, Gentier 48E, Gentier 48E.

Do not use the kit or any component after the expiry date. Do not substitute reagents from

different kit batches or reagents from other commercially available kits.

The PCR amplification stage is exceedingly susceptible to cross-contamination. The work flow of all assay equipment and consumables must be from pre-amplification to post-

To help prevent contamination change gloves frequently and use different sterile, nuclease free pipette tips for each sample and reagent.
The Positive Control contains an RNA oligo of the native detection target so keep well away

from the other reagents to prevent contamination which could lead to false positive results. Never open the post-amplification tubes. Instead, wrap and seal them up and discard the tubes along with unused reagent, assay consumables and assay waste according to local regulations.

Storage and Stability:

The kit is to be stored refrigerated at 2 - 8°C and protected against exposure to light. The kit is stable up to the expiry date printed on the outer label. The kit should be transported in a polystyrene lined box containing ice packs ($2-28^{\circ}$ C) and delivered within 14 days.

Sample Collection and Preparation:

The COVID-19 Lyophilised Nucleic Acid Kit can be performed using viral RNA extracted from

upper and lower respiratory tract samples.
Upper respiratory tract samples should be collected via nasopharyngeal or pharyngeal wash

or swabs by recommended technique. Lower respiratory tract samples should be collected via bronchoalveolar lavage, tracheal

extract or deep cough sputum by recommended technique. Swab samples should be collected only using swabs with a synthetic tip and aluminium or plastic shaft. Do not use swabs with calcium alginate or cotton tips or with wooden shafts

Ideally RNA extraction from the samples should be performed immediately. Alternatively, the samples may be stored up to 24 hours at 2 – 8oC or for longer at -70oC. Avoid repeat freeze-thawing.

Viral RNA extraction reagents are not included in this kit. Extract RNA following the Instructions of Use for the viral RNA extraction kit of choice.

- A Detection Remove freeze dried Buffer Mix tube(s) and Reagent Solution tube(s) from the kit (see below to calculate number of tubes required). Reconstitute the Buffer Mix tube by adding 900 µl Reagent Solution to form the Working Mix. Mix by repeatedly withdrawing and expelling the Mix using a pipette and tip (at least 10x) then centrifuge briefly. A volume of 45 µl/test

- Every PCR run must include one Positive Control and one Negative Control so include these in volume required calculations. Label tubes for these and for each sample being run.
- Pipette 45 µl Working Mix into each labelled tube.
- Bring RNA samples to room temperature and take out the Positive Control and Negative Control from the kit. Reconstitute the Positive Control (Buffer PC) tube with 50 μl Reagent Solution.
- Add 5 µl Negative Control (Buffer NC), 5 µl RNA sample, or 5 µl Positive Control (preferably in that order) to assigned tubes using separate pipette tips. Cap the tubes.

 Briefly centrifuge the tubes to force all volume to the base of the tubes.

- Programme the analyser according to the following cycling parameters:

Stage	Cycles	Temp (°C)	Time	Data
Reverse transcription	1	48	15 min	N/A
Initial denaturation	1	93	2 min	N/A
Cycle reaction	45	93	15 sec	N/A
		58	40 sec	FAM, HEX/VIC, ROX
Device Cooling	1	30	30 sec	

- Initiate the PCR run immediately
- When the run is complete analyse the data, see below.

Baseline setting: Adaptive baseline.
Threshold setting: The Threshold line should be set just above the peak of the normal negative control amplification curve (irregular noise line).

Analysis of Results:

Assay Validation

- Check the results for the Controls first:

Control Result	ROX (Internal	FAM (ORF1ab	HEX (N gene)
	Control, β-actin)	gene)	
Negative Control	No Ct or Ct = 0	No Ct or Ct = 0	No Ct or Ct = 0
Positive Control	Ct < 40, with good	Ct < 42	Ct < 42
	amplification curve		

- If Controls meet all the validation criteria above, proceed to sample analysis

Sample Analysis

Sample	ROX (Internal	FAM (ORF1ab	HEX (N gene)
Result	Control, β-actin)	gene)	
Negative	Ct < 40, with good	No Ct or Ct = 0	No Ct or Ct = 0
Positive	amplification curve	Ct < 42	Ct < 42

If the result for FAM or HEX fall within the range 42 ≤ Ct < 45 the experiment must be repeated. If Ct <45 with a good amplification curve, the sample may be considered to

A sample is considered positive when ORF1ab and N of 2019-nCoV are both positive. Each sample defined positive using this kit should be repeated using a different detection method using re-extracted RNA.

Performance Characteristics:

Sensitivity limit: 300 copies/ml. Coincidence rate of Positive reference: 100% Coincidence rate of Negative reference: 100%.

Precision: CV% < 5%

Limitations of the Test:

The COVID-19 Lyophilised Nucleic Acid Kit is only to be used by laboratory personnel trained

The results obtained with this assay should not be used as the sole criterion for diagnosis of COVID-19 infection, but be used in conjunction with other diagnostic procedures and clinical

Results will only be accurate if samples are properly processed and correctly transported and stored. If the RNA is degraded the ability of this kit to detect COVID-19 will be compromised.

References:

- World Health Organisation Statement regarding cluster of pneumonia cases in
- Wuhan, China: 9 January 2020.

 Xintian Xu et al. Evolution of the coronavirus from the ongoing Wuhan outbrek and modelling of its spike protein for risk of human transmission. Published
- online 21 Jan 2020 World Health C Organisation. Coronavirus. www. who.int/health-topics /coronavirus.

Glossary of Symbols:

REF	Catalogue number	\mathcal{A}	Temperature limitation
Consult instructions for use		LOT	Batch code
IVD	In vitro diagnostic medical device	X	Use by
***	Manufacturer		Do not reuse
类	Keep away from sunlight		



Prestige Diagnostics UK Ltd, 40 Ballymena Business Centre, Galgorm, Co. Antrim, BT42 1FL, United Kingdom. Tel: +44 (0) 28 25642100